

Evolution of Phosphagen Kinase (III). Amino Acid Sequence of Arginine Kinase from the Shrimp *Penaeus japonicus*

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ABSTRACT—The amino acid sequence of arginine kinase (AK) from the shrimp *Penaeus japonicus* has been determined chemically. It consists of 355 amino acid residues, and has a calculated molecular mass of 40,018 Da. The amino acid sequence of *Penaeus* AK showed 91% and 51% identity, respectively, with those of AKs from the lobster *Homarus vulgaris* and the abalone *Nordotis madaka*. It also showed significant homology (39–43%) with vertebrate or invertebrate creatine kinases and annelid glycoyamine kinase, suggesting that these enzymes evolved from a common origin.

INTRODUCTION

Phosphagen kinases (PKs) are the enzymes that catalyze the reversible transfer of the high energy phosphoryl group of ATP to the naturally occurring guanidines, and play a key role to interconnect energy production and utilization in animals [9]. In vertebrates, the only phosphagen kinase is creatine kinase (CK), but in invertebrates, at least five phosphagen kinases, arginine kinase (AK), glycoyamine kinase (GK), taurocyamine kinase (TK), lombricine kinase (LK) and CK, have been identified by partial or complete sequencing [1–3, 15, 18, 20]. Moreover, the presences of hypotaurocyamine kinase (HTK), opheline kinase (OK) and thalassemine kinase (ThalK) are proposed by their enzyme activity [11, 19]. The homologous amino acid sequences of about 15 residues around the putative active site of these enzymes suggest that they have evolved from a common origin [1], and thus provide an excellent model system to elucidate how enzymes developed the recognition site for substrate during evolution.

AK is the phosphagen kinase that is most widely distributed in animals. Very recently, two cDNA-derived amino acid sequences of AKs from the lobster *Homarus vulgaris* [3] and the abalone *Nordotis madaka* [18] have been determined. Here we report the primary structure of AK from the shrimp *Penaeus japonicus*, to be sequenced chemically. A preliminary account of this work has been presented [13].

MATERIALS AND METHODS

AK was isolated from the tail muscle of *Penaeus japonicus* according to our previous method [12].

The protein (50 nmoles) was carboxymethylated and cleaved with CNBr in 70% formic acid. Larger CNBr peptides were digested further with lysyl endopeptidase, chymotrypsin, *S. aureus* V8 protease and pepsin. To obtain overlap peptides, the protein was also digested with lysyl or arginyl endopeptidases. The diges-

tion conditions are the same as described previously [16]. The digested products were purified on a reverse-phase column (Cosmosil 5C₁₈-300, 2.5×150 mm) with a linear gradient of 0–80% acetonitrile in 0.1% trifluoroacetic acid (TFA) at a flow rate of 1 ml/min. Some peptides were purified further by rechromatography. Amino acid analyses and the manual Edman sequencing of the peptides were done with our standard methods [16]. The N-terminal peptide CN1C1 was digested with acylamino acid releasing enzyme (0.025U, Takara) in 5 mM phosphate buffer (pH 7.2) containing 1 mM 2-mercaptoethanol at 37°C for 2 hr, before sequencing.

RESULTS AND DISCUSSION

Fig. 1 shows the elution profile of CNBr peptides of *Penaeus* AK on reverse-phase chromatography. Most of the CNBr peptides were separated successfully, and the larger peptides were digested further with several enzymes. Two small CNBr peptides, Gln-Met at positions 233–234 and C-terminal Glu-Lys-Glu-Met, were not recovered. The overlap of CNBr peptides was obtained with the peptides derived from lysyl or arginyl endopeptidase digestions of the whole protein. Amino acid compositions of the CNBr peptides and the whole protein are given in Table 1. The strategy to establish the complete amino acid sequence is shown in Fig. 2. The Gly-Arg bond at position 206–207, Arg-Ala at 279–280, Arg-Gly at 308–309 and Lys-Arg at 327–328 were unusually cleaved with *S. aureus* V8 protease. The sequence is supported by at least two amino acids overlap, and the C-terminal half of 129 residues was also confirmed by the cDNA sequencing [17]. *Penaeus* AK begins with the blocked Val, is composed of total 355 amino acid residues and the molecular mass was calculated to be 40,018 Da.

So far, all the amino acid sequences of phosphagen kinases were determined by their cDNA sequencing, and therefore this work is the first example to be sequenced chemically.

Amino acid sequence of *Penaeus* AK was aligned with *Homarus* AK, *Schistosoma* (trematode) PK domain 1, *Neanthes* (annelid) GK, *Nordotis* (mollusc) AK, sea urchin CK domains 2 and 3 and chicken three CK isoforms (muscle, brain and mitochondrial types), with the algorithm of Feng &

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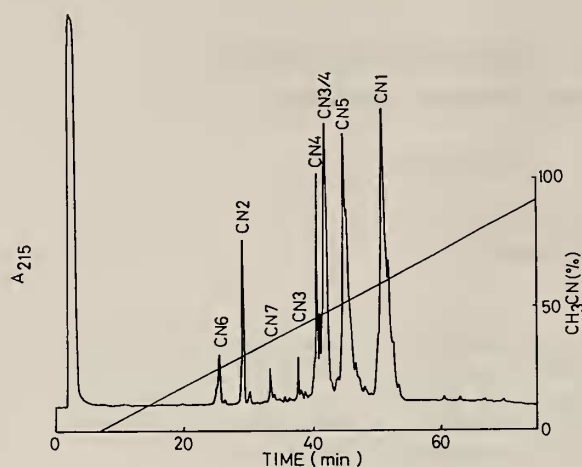


Fig. 1. Elution profile of CNBr peptides of *Penaeus* AK on reverse-phase chromatography. The column (Cosmosil 5C₁₈-300, 2.5 × 150 mm) was eluted with a linear gradient of 0–80% acetonitrile in 0.1% TFA at a flow rate of 1 ml/min.

Doolittle [5], in Fig. 3. It is noted that sea urchin CK has an unusual three-domain structure which may be resulted from a gene triplication [20] and *Schistosoma* PK has a two-domain structure, of which the second domain lacks the C-terminal 50 residues [15]. *Penaeus* AK is aligned with *Homarus* AK without any insertions or deletions. Furthermore, their sequences are characterized by a unique deletion at positions 117–118 and an insertion at position 311 in Fig. 3. In the alignment, there are 68 amino acid residues (indicated by asterisks) conserved in all of the phosphagen kinases.

All of the phosphagen kinases can be inactivated partial-

ly or completely under the chemical modification with thiol-specific reagents [5]. The reactive cysteine, that would be located near or in the center of the putative active site, was identified as Cys-286 (see Fig. 3). Recent site-directed mutagenesis study shows that the active cysteine is necessary to confer conformational changes upon substrate binding, but is not essential for catalysis [6].

The percent identity between the 10 amino acid sequences of phosphagen kinases is shown in Table 2. The sequence of *Penaeus* AK showed 91% and 51% identity, respectively, with those of *Homarus* and *Nordotis* AKs. It also showed significant homology (39–43%) with vertebrate or invertebrate CKs and *Neanthes* GK. These sequence homologies would be enough to conclude that CK, GK and AK are derived from a common origin.

A phylogenetic tree was constructed from the sequence alignment in Fig. 3 with the algorithm of Feng & Doolittle [5] (Fig. 4). The same topology was also obtained with the protein parsimonious algorithm using the program *Protpars* in the PHYLIP package ver 3.5c [4]. The tree separated phosphagen kinases into two clusters, a cluster containing vertebrate and invertebrate CKs and invertebrate GK and a cluster containing three AKs and *Schistosoma* PK. The branching pattern clearly shows that CK and GK must have evolved from a common ancestor [18]. The phylogenetic position of *Schistosoma* PK is noted. Our tree placed the PK near the cluster of AKs (Fig. 4). In fact, *Schistosoma* PK has the highest sequence homology (46–52%) with AKs (see Table 2). Moreover, *Schistosoma* PK shares the sequence characteristics with invertebrate AKs: deletions at posi-

TABLE 1. Amino acid compositions of *Penaeus* AK and its CNBr peptides

	whole	CN1	CN2	CN3	CN3/4	CN4	CN5	CN6	CN7
Asx	37.8(36)	15.4(16)	1.0(1)		8.8(9)	8.6(9)	9.3(9)		1.1(1)
Thr	16.9(19)	5.4(6)	1.0(1)	3.0(3)	3.6(4)	1.0(1)	6.2(7)	1.1(1)	
Ser	15.8(19)	7.1(9)		3.4(4)	5.2(6)	1.9(2)	3.4(4)		
Glx	11.0(11)	11.0(11)	4.1(4)	4.5(4)	11.9(11)	7.5(7)	10.2(9)	3.6(3)	2.3(2)
Pro	10.6(12)	4.8(5)	1.9(2)	0.8(1)	1.9(2)	0.9(1)	2.9(3)		
Gly	31.2(29)	9.5(10)	1.0(1)	3.3(3)	6.2(6)	3.3(3)	9.9(10)	1.1(1)	1.1(1)
Ala	22.6(22)	9.7(10)	1.0(1)		4.0(4)	3.9(4)	6.2(6)	1.1(1)	
Cys	5.0(5)	1.5(2)	0.9(1)		0.8(1)	0.9(1)	1.0(1)		
Val	22.6(22)	9.1(11)		1.2(1)	4.1(4)	3.1(3)	6.0(6)	1.3(1)	
Met	9.7(9)	+ (1)	+ (1)	+ (1)	+ (2)	+ (1)	+ (1)	+ (1)	+ (1)
Ile	15.8(17)	3.8(6)			2.7(4)	2.8(4)	4.5(5)		1.6(2)
Leu	37.0(35)	12.5(13)	1.1(1)	4.3(4)	8.9(9)	4.9(5)	9.7(9)	1.3(1)	1.8(2)
Tyr	11.9(12)	4.4(5)	1.8(2)	0.5(1)	2.4(3)	1.3(2)	2.0(2)		
Phe	18.9(18)	6.4(7)	1.0(1)	1.1(1)	5.1(5)	3.9(4)	4.2(4)	1.3(1)	
Lys	26.8(29)	12.0(13)	1.0(1)	2.0(2)	6.0(6)	4.0(4)	6.0(6)	1.0(1)	1.0(1)
His	10.1(9)	1.7(2)			2.9(3)	2.7(3)	3.8(4)		
Arg	15.6(17)	2.7(3)			3.7(4)	3.6(4)	9.5(10)		
Trp	+ (2)				+ (2)	+ (2)			
Total	(355)	130	17	25	85	70	96	11	10
Position		1–130	131–147	148–172	148–232	173–232	235–330	331–341	342–351
Yield(%)		57.9	42.3	9.0	35.4	15.6	41.0	34.5	46.7

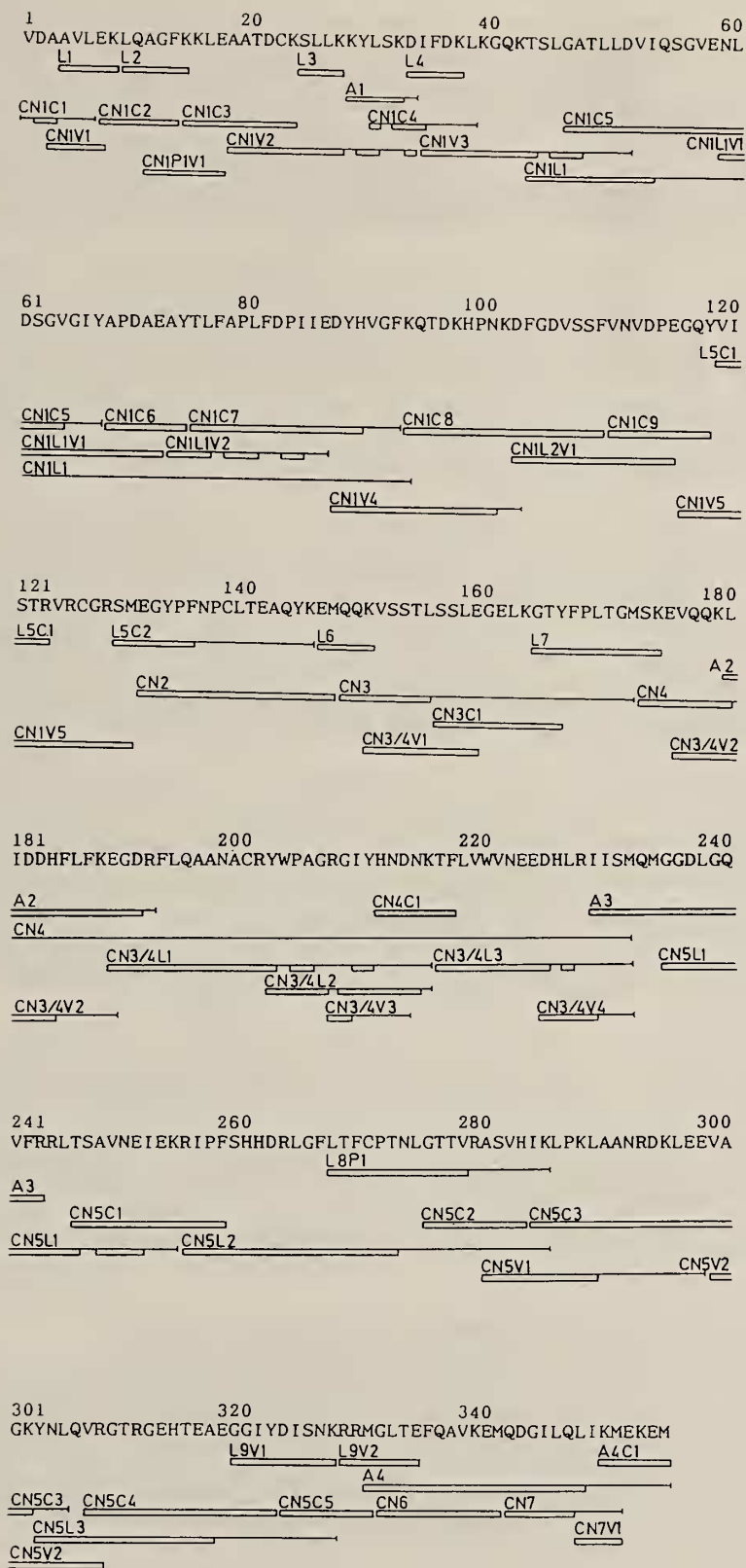


FIG. 2. Summary of data to establish the amino acid sequences of *Penaeus* AK. The sequence was determined by manual Edman degradation (□). Key; CN, CNBr; L, lysyl endopeptidase; C, chymotrypsin; V, *S. aureus* V8 protease; P, pepsin; A, arginyl endopeptidase.

25 50

AK shrimp VDAVLEKQ AGFKLEAATDCKSLKKYL SKDIFDKLKGQKTSLGATLLDVIQSGVENLD
 AK lobster MADAATIAKLE EGFKLEAATDCKSLKKYL SKDIFDSLKAKKTSLGATLLDVIQSGVENLD
 AK abalone MLAMASVE ELWAKLDGAADCKSLKNNLTKERYEALKDKKTKFGGTLADCI RSGCLNLD
 PK Schis.1 MQVESLQ NLQAKIRNDRNHS LTKKYL TDDIVKKYQATKTSLGATLLDVIQSGVENLD
 CK-M chick PFSSTHNKHLKFSAE EFPDL SKHNNHMAKVL TP ELYKRLRDKETPSGFTLDDV IOTGVDNPGH
 CK-B chick PFSNSHNLKMKYSVDDEY PDL SVHNNHMAKVL TLDLYKRLRDRQTS SSGFTLDDV IOTGVDNPGH
 CK sea ur2 YPDL SKHNNH LAHCL TYDIWKS LKDKKTPSGFTLDDV IOTGVDNPGH
 CK sea ur3 YPDFSLHNNWMSK CMTE EYKLCNLKTKGGV TLNDCIOTGIDNPGH
 CK-Mt chic TVHEKRKL FPPSADY PDLRKHNNCMAECLTPAIYAKLRDKLTPNGYSLDQCIOTGVDNPGH
 GK ma.worm MFKDYSREKF AKENFPDL SKHNNVMASHLTYELYEKYWDKVPNGVTLDDKCIOTGVDNPGH

75 100 125

AK shrimp SGVGIYAPDAEAYTLFAPLFDPIIEDYHVGFKQTDKHPNKDFGDVSS FVNVDPGEQYVI
 AK lobster SGVGIYAPDAEAYSLFAPLFDPIIEDYHKGFKQTDKHPAKDFGDVSK FINVDPEGT FVI
 AK abalone SGVGIYACDPDAYTVFADVLDAVIKEYHKV PELKHPEPEMGDLKLNFGDLPDPSGEYIV
 PK Schis.1 ALLPR SCDLNAYETFRDFDAVIADYHKVPDGKIQHPKSNFGDLKSLSFTDLNTYGNL VV
 CK-M chick PFI MTVGCVAGDEESYEVFKDLFDPVIO DRHGGYK TDKHRTDLNHNELKGGDDLD P KYVL
 CK-B chick PFI MTVGCVAGDEESYEVFKELFDPVIEDRHGGYK TDEHKTDLNADNLQGGDDLD P NYVL
 CK sea ur2 PHI MTVGMVAGDEESYDV FADIFDPVIDARHGGYK DAVHVTNINHADL KGGDNLDP KYVL
 CK sea ur3 PYI MTVGLVAGDEECYEVFAPLFDPVISARHGGYAL DAKHPTLNAAELKGGDDLD P EFVL
 CK-Mt chic PFI KTVGMVAGDEESYEVFAEIFDPVIKARHNGYDPR TMKHHTDL DASKITHG QFDE RYVL
 GK ma.worm KFYGKKTGCVFGDEHSYETFKDFDRVIEEIH FKPEDVHPATDLDETKLVGG VFDE KYVK

150 175

AK shrimp STRVRCGRSMEGYFPNCLTEAQYKEMQOKVSS TSSLEGE LKGTFFLTGMSKEVQOKLIDDFH
 AK lobster STRVRCGRSMEGYFPNCLTEAQYKEME EKVSS TSSLEGE LKGSYFPLTGMTKEVQOKLIDDFH
 AK abalone STRVRVGRSHDSYGFPPVLTKQERL KMEEDTKAAFEKFSGELAGKYFPLEGMSKEDQKQMTEDHF
 PK Schis.1 STRVRLGRTVGEGFGPRTLTKRIELENKISTALHNLSGEYEGTYPLTGCGRQNTSKRHHF
 CK-M chick SSVRVTGRS IKGYS LPPHCSRGERRAVEKLSVEALNSLEGEFKGRYYPLKAMTEQEQQQLTDDHF
 CK-B chick SSVRVTGRSIRGFC LPPHCSRGERRAIEKLSVEALGSLGGDLKGGYALRNMTDAEQQLIDDFH
 CK sea ur2 SCRVRTGRS IIGYSLPPHCVEERA AVETIIGALDKFDGDLQGGYPL EGMSTDETOTQLIDDFH
 CK sea ur3 SCRVRTGRSIRGLALPPCCTRAERA EVEKITTEALSTL SGPLKGGYPLTGMTDEEQEKLIEDHF
 CK-Mt chic SSVRVTGRSIRGLS LPPACSRARERVENVVV TALAGLKGDL SGKYSSLT NMSERDQQQLIDDFH
 GK ma.worm SCRIRCGRSVRGVC LPPAMSAERRLVEKVVSNALGGLKEDLAGKYFPLTMMNDKMEALIEDHF

200 225 250

AK shrimp LF KEGDRFLQAANACRYWPA GRGIYHNDNK TFLVWVNEEDHLRIISMQMGDDLQGVFRRLTSAV
 AK lobster LF KEGDRFLQAANACRYWPA GRGIYHNDNK TFLVWCNEEDHLRIISMQMGDDLQGVYRRLVSAV
 AK abalone LF KDDDRFLRDAGGYNDWCSGRGIFNTAKN FLVWVNEEDHLRLISMQGGDLAAVYKRLVVAI
 PK Schis.1 LF RNDNVL RDAAGYIDWPTGRGIFINKQKFLVWINEEDHIRVISMQGRDLIAVYKRLADAI
 CK-M chick LFDKPV SPLLLASGMARDWPDARGIWHNDNK TFLVWVNEEDHLRVISMQGGNMKEVFRFCTGL
 CK-B chick LFDKPV SPLLLASGMARDWPDARGIWHNDNK TFLVWINEEDHLRVISMQGGNMKEVFRFCTGL
 CK sea ur2 LFDKPV SPLLLAARMHRDWPQGRGIWHNENK NFLVWVNEEDHIRVISMKDGMMRAVFKRCEGL
 CK sea ur3 LFDKPV SPLLLCANMARDWPQGRGIWHNDEKNFLVWVNEEDHTRVISMKSGNMKRVFERFCGL
 CK-Mt chic LFDKPV SPLLLTCAGMARDWPDARGIWHNDNK TFLVWINEEDHTRVISMKSGNMKRVFERFCRGL
 GK ma.worm LFEKPTGALLTTSGCARDWPDARGIWHNNGKNFLVWINEEDHIRIISMQGGDMRAVFSRFRGL

275 300 325

AK shrimp NEIE KR IPFSHHDRLGFLTFcPTNLGTTVRASVHIKLPKLAANRDKLEEVAGKYNLQVRG
 AK lobster NDIE KR VPFSHHDRLGFLTFcPTNLGTTVRASVHIKLPKLAANREKLEEVAAKFSLQVRG
 AK abalone NTMT ASGLSFAKRDGLGFLTFcPSNLGTALRASVHMKIPNLAASPE FKSFCNLDNLQARG
 PK Schis.1 QELS KS LKFAFNDR LGFITFcPSNLGTTLRASVHAKIPMLASLPN FKEICEKHGIQPRG
 CK-M chick KKIEEIFKKA GHPFMWTEHLGYILTcPSNLGTGLRGVHVKLPKLSQHPK FEEILHRLRLQKRG
 CK-B chick TQIETLFKSKNYEFMWNPHLGYILTcPSNLGTGLRAGVHIKLPNLKHEK FGEVILKRLRLQKRG
 CK sea ur2 QKFEQMIKKDGKEFMWNKHLGYVLTcPSNLGTGLRAGVHVKLPNLSKYPR FQDILRALRLQKRG
 CK sea ur3 KKVEDSIKSKGYQFMWNEHLGYVLTcPSNLGTGLRAGVHVKLPNLSQOKI FDSILDHMLRLQKRG
 CK-Mt chic KEVERLIKERGWEFMWNERLGYVLTcPSNLGTGLRAGVHVKLPRLSKDPR FPKILENRLQKRG
 GK ma.worm TEVERLMKEKGYELMRNERLGYICTcPTNLGTTVRASVHLRLANLEKDKR FDFFLAKLRLGKRG

350 375

AK shrimp TRGEHTEAEGGIYDISNKRRMGLTEFQAVKEMQDGILQLIKMEKEM
 AK lobster TRGEHTEAEGGIYDISNKRRMGLTEFQAVKEMQDGILELIKIEKEM
 AK abalone IHGEHTESVGGYDLSNKRRRLGLTEYQAVEEMRVGVEACLAKELAAAK
 PK Schis.1 THGEHTESVGGIYDLSNKRRRLGLTELDV TEMHSGVRALLEEVLMEYKNGAPEGV
 CK-M chick TGGVDTAAV GAVFDISNADRLGFSEVEQVMVVDGVKLMVEMEKLEQNPIDDMIPAQK
 CK-B chick TGGVDTAAVGGVFDVSNADRLGFSEVELVQMVVDGVKLLIEMEKRLKQGSIDDLMPAQK
 CK sea ur2 TGGVDTASTDGTFDISNLDRLGSSEVQVQVVDVGVVLLVQMEKKLEKGEDIFDL PQQCRPKPP
 CK sea ur3 TGGVDTASTDGTFDISNDRIGFSEVHLVQQLVDGVKLLVNLEKALMKGEDINSLLPEKLRDSS
 CK-Mt chic TGGVDTAAVADVDISNLDRLMGRSEVELVQIVIDGVNVLVDCEKLEKQDQKVPPLPOFGRK
 GK ma.worm TGGESSLAEDSTYDISNLDRLGKSERELVQVLDGVNVLIEADKRLEAGKPIDDLTPRLNSSTGT

400

AK shrimp
 AK lobster
 AK abalone
 PK Schis.1
 CK-M chick

CK-B chick
 CK sea ur2 IKPFSYD
 CK sea ur3
 CK-Mt chic
 GK ma.worm SISATASRHMTL

FIG. 3. Alignment of the amino acid sequences of 10 phosphagen kinases. This alignment was obtained with the algorithm of Feng & Doolittle [5]. Invariable residues are indicated by asterisks. The reactive cysteine is shown by \$. References; CK-M chick (chicken muscle isoform) [10, 14]; CK-B chick (chicken brain isoform) [7]; CK sea ur2 and 3 (domains 2 and 3 of sea urchin) [20]; CK-Mt chic (chicken mitochondrial isoform) [8]; GK ma.worm (*Neanthes*) [18]; AK shrimp (*Penaeus*) (this work); AK abalone (*Nordotis*) [18]; AK lobster (*Homarus*) [3]; PK Schis. 1 (domain 1 of *Schistosoma*) [15].

TABLE 2. Percent identity between the sequences of phosphagen kinases

	AK lob	AK aba	PK Sch	CK-M	CK-B	CK ur2	CK ur3	CK-Mt	GK wor
Ak shr	91.0	50.9	46.3	42.5	43.0	42.3	41.7	39.8	38.5
AK lob		52.6	45.7	41.8	41.8	41.7	41.7	39.3	38.8
AK aba			51.8	38.3	36.9	43.3	39.8	37.3	37.1
PK Sch				34.4	35.8	36.8	36.2	35.9	34.6
CK-M					80.3	68.2	65.2	67.2	53.2
CK-B						65.8	65.8	66.7	56.2
CK ur2							68.9	65.5	51.1
CK ur3								64.1	51.0
CK-MT									57.4

Abbreviations: shr, *Penaeus*; lob, *Homarus*; aba, *Nordotis*; Sch, domain 1 of *Schistosoma*; CK-M, chicken muscle isoform; CK-B, chicken brain isoform; ur2 and ur3, domains 2 and 3 of sea urchi; CK-Mt, chicken mitochondrial isoform; wor, *Neanthes*.

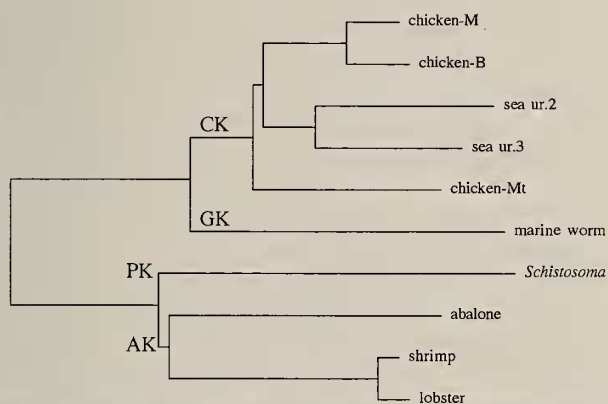


FIG. 4. A phylogenetic tree constructed from 10 sequences of phosphagen kinases aligned in Fig. 2. The tree was obtained with the program of Feng & Doolittle [5].

tions 13, 65-69, 198 and 265-268 in Fig. 3. Stein *et al.* [15] assigned tentatively *Schistosoma* PK as CK, based on the very weak, but reproducible CK activity in the crude extracts. However, since the enzyme activities of CK, GK, AK are strictly specific and those of TK, HTK, LK, OK and ThalK are more or less interspecific [1, 19], it is very likely that *Schistosoma* PK belongs to a member of the latter group. Dumas & Camonis [3] also suggest this possibility, based on the higher % identity between *Schistosoma* PK and lobster AK.

The evolutionary origin of phosphagen kinases is of

primary concern. Of the phosphagen kinases, AK is most widely distributed in animals. However the wide distribution does not imply that AK is closer to an ancestral phosphagen kinase. To solve this problem, we are planning to analyze the phosphagen kinases from more primitive animals, such as sea anemones and protozoa.

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